



CLINICAL AND  
LABORATORY  
STANDARDS  
INSTITUTE

34th Edition

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# CLSI M100™

## Performance Standards for Antimicrobial Susceptibility Testing

Sample

CLSI M100 includes updated tables for the Clinical and Laboratory Standards Institute antimicrobial susceptibility testing standards CLSI M02, M07, and M11.

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A CLSI supplement for global application.

# Performance Standards for Antimicrobial Susceptibility Testing

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## Abstract

The data in the tables are valid only if the methodologies in CLSI M02,<sup>1</sup> M07,<sup>2</sup> and M11<sup>3</sup> are followed. These standards contain information about disk diffusion (CLSI M02<sup>1</sup>) and dilution (CLSI M07<sup>2</sup> and CLSI M11<sup>3</sup>) test procedures for aerobic and anaerobic bacteria. Clinicians depend heavily on information from the microbiology laboratory for treating their seriously ill patients. The clinical importance of antimicrobial susceptibility test results demands that these tests be performed under optimal conditions and that laboratories have the capability to provide results for the newest antimicrobial agents. The tables presented in CLSI M100 represent the most current information for drug selection, interpretation, and quality control using the procedures standardized in CLSI M02,<sup>1</sup> M07,<sup>2</sup> and M11.<sup>3</sup> Users should replace previously published tables with these new tables. Changes in the tables since the previous edition appear in boldface type.

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# Contents

- Abstract ..... i
- Committee Membership ..... iii
- Overview of Changes ..... xii
- CLSI Breakpoint Additions Since 2010 ..... xx
- CLSI Breakpoint Revisions Since 2010 ..... xxiii
- CLSI Archived Resources ..... xxvii
- Summary of CLSI Processes for Establishing Breakpoints and QC Ranges ..... xxviii
- CLSI Reference Methods vs Commercial Methods and CLSI vs US Food and Drug Administration Breakpoints ..... xxix
- CLSI Subcommittee on Antimicrobial Susceptibility Testing Mission Statement ..... xxx
- Instructions for Use of Tables ..... 1
- References ..... 20
- Introduction to Tables 1A–1J. Antimicrobial Agents That Should Be Considered for Testing and Reporting by Microbiology Laboratories ..... 22
- Table 1A-1. Enterobacterales (excluding *Salmonella/Shigella*) ..... 24
- Table 1A-2. *Salmonella* and *Shigella* spp. .... 26
- Table 1B-1. *Pseudomonas aeruginosa* ..... 28
- Table 1B-2. *Acinetobacter* spp. .... 30
- Table 1B-3. *Burkholderia cepacia* complex ..... 32
- Table 1B-4. *Stenotrophomonas maltophilia* ..... 34
- Table 1B-5. Other Non-Enterobacterales ..... 36
- Table 1C. *Staphylococcus* spp. .... 38
- Table 1D. *Enterococcus* spp. .... 40
- Table 1E. *Haemophilus influenzae* and *Haemophilus parainfluenzae* ..... 42
- Table 1F. *Neisseria gonorrhoeae* ..... 44

## Contents (Continued)

Table 1G. <i>Streptococcus pneumoniae</i> .....	46
Table 1H-1. <i>Streptococcus</i> spp. $\beta$ -Hemolytic Group.....	48
Table 1H-2. <i>Streptococcus</i> spp. Viridans Group.....	50
Table 1I. <i>Neisseria meningitidis</i> .....	52
Table 1J. Anaerobes.....	54
Introduction to Tables 2A–2J. Zone Diameter and MIC Breakpoints.....	56
Table 2A-1. Zone Diameter and MIC Breakpoints for Enterobacterales (excluding <i>Salmonella/Shigella</i> ).....	58
Table 2A-2. Zone Diameter and MIC Breakpoints for <i>Salmonella</i> and <i>Shigella</i> spp.....	70
Table 2B-1. Zone Diameter and MIC Breakpoints for <i>Pseudomonas aeruginosa</i> .....	74
Table 2B-2. Zone Diameter and MIC Breakpoints for <i>Acinetobacter</i> spp.....	80
Table 2B-3. MIC Breakpoints for <i>Burkholderia cepacia</i> complex.....	86
Table 2B-4. Zone Diameter and MIC Breakpoints for <i>Stenotrophomonas maltophilia</i> .....	88
Table 2B-5. MIC Breakpoints for Other Non-Enterobacterales.....	92
Table 2C. Zone Diameter and MIC Breakpoints for <i>Staphylococcus</i> spp.....	96
Table 2D. Zone Diameter and MIC Breakpoints for <i>Enterococcus</i> spp.....	106
Table 2E. Zone Diameter and MIC Breakpoints for <i>Haemophilus influenzae</i> and <i>Haemophilus parainfluenzae</i> .....	112
Table 2F. Zone Diameter and MIC Breakpoints for <i>Neisseria gonorrhoeae</i> .....	118
Table 2G. Zone Diameter and MIC Breakpoints for <i>Streptococcus pneumoniae</i> .....	122
Table 2H-1. Zone Diameter and MIC Breakpoints for <i>Streptococcus</i> spp. $\beta$ -Hemolytic Group.....	128
Table 2H-2. Zone Diameter and MIC Breakpoints for <i>Streptococcus</i> spp. Viridans Group.....	134
Table 2I. Zone Diameter and MIC Breakpoints for <i>Neisseria meningitidis</i> .....	138

## Contents (Continued)

Table 2J. MIC Breakpoints for Anaerobes .....	142
Introduction to Table 2 Dosages. Antimicrobial Agent Dosage Regimens Used to Establish Susceptible or Susceptible-Dose Dependent Breakpoints .....	146
Table 2 Dosages. Antimicrobial Agent Dosage Regimens Used to Establish Susceptible or Susceptible-Dose Dependent Breakpoints .....	148
Table 3A. Tests for Extended-Spectrum $\beta$ -Lactamases in <i>Klebsiella pneumoniae</i> , <i>Klebsiella oxytoca</i> , <i>Escherichia coli</i> , and <i>Proteus mirabilis</i> .....	154
Introduction to Tables 3B and 3C. Tests for Carbapenemases in Enterobacterales and <i>Pseudomonas aeruginosa</i> .....	158
Table 3B. CarbaNP Test for Suspected Carbapenemase Production in Enterobacterales and <i>Pseudomonas aeruginosa</i> .....	160
Table 3C. Modified Carbapenem Inactivation Methods for Suspected Carbapenemase Production in Enterobacterales and <i>Pseudomonas aeruginosa</i> .....	168
Table 3D. Aztreonam Plus Ceftazidime-Avibactam Broth Disk Elution Method .....	182
Table 3E. Tests for Colistin Resistance for Enterobacterales and <i>Pseudomonas aeruginosa</i> .....	192
Table 3F-1. Test for Performing Disk Diffusion Directly From Positive Blood Culture Broth .....	198
Table 3F-2. Zone Diameter Disk Diffusion Breakpoints for Enterobacterales Direct From Blood Culture .....	202
Table 3F-3. Zone Diameter Disk Diffusion Breakpoints for <i>Pseudomonas aeruginosa</i> Direct From Blood Culture .....	204
Table 3F-4. Zone Diameter Disk Diffusion Breakpoints for <i>Acinetobacter</i> spp. Direct From Blood Culture .....	206
Table 3G. Tests for Detecting $\beta$ -Lactamase Production in <i>Staphylococcus</i> spp. ....	208
Table 3H. Oxacillin Salt Agar Test for Detecting Methicillin (Oxacillin) Resistance in <i>Staphylococcus aureus</i> .....	212
Table 3I. Vancomycin Agar Screen for <i>Staphylococcus aureus</i> and <i>Enterococcus</i> spp. ....	214
Table 3J. Tests for Detecting Inducible Clindamycin Resistance in <i>Staphylococcus</i> spp., <i>Streptococcus pneumoniae</i> , and <i>Streptococcus</i> spp. $\beta$ -Hemolytic Group .....	216
Table 3K. Test for Detecting High-Level Mupirocin Resistance in <i>Staphylococcus aureus</i> .....	220
Table 3L. Test for Detecting High-Level Aminoglycoside Resistance in <i>Enterococcus</i> spp. (including disk diffusion) .....	222

## Contents (Continued)

Table 4A-1. Disk Diffusion QC Ranges for Nonfastidious Organisms and Antimicrobial Agents Excluding $\beta$ -Lactam Combination Agents .....	226
Table 4A-2. Disk Diffusion QC Ranges for Nonfastidious Organisms and $\beta$ -Lactam Combination Agents .....	232
Table 4B. Disk Diffusion QC Ranges for Fastidious Organisms .....	236
Table 4C. Disk Diffusion Reference Guide to QC Frequency .....	240
Table 4D. Disk Diffusion Troubleshooting Guide .....	242
Table 5A-1. MIC QC Ranges for Nonfastidious Organisms and Antimicrobial Agents Excluding $\beta$ -Lactam Combination Agents .....	248
Table 5A-2. MIC QC Ranges for Nonfastidious Organisms and $\beta$ -Lactam Combination Agents .....	258
Table 5B. MIC QC Ranges for Fastidious Organisms (Broth Dilution Methods) .....	264
Table 5C. MIC QC Ranges for <i>Neisseria gonorrhoeae</i> (Agar Dilution Method) .....	270
Table 5D. MIC QC Ranges for Anaerobes (Agar Dilution Method) .....	272
Table 5E. MIC QC Ranges for Anaerobes (Broth Microdilution Method) .....	276
Table 5F. MIC Reference Guide to QC Frequency .....	278
Table 5G. MIC Troubleshooting Guide .....	280
Table 6A. Solvents and Diluents for Preparing Stock Solutions of Antimicrobial Agents .....	288
Table 6B. Preparing Stock Solutions for Antimicrobial Agents Provided With Activity Expressed as Units .....	296
Table 6C. Preparing Solutions and Media Containing Combinations of Antimicrobial Agents .....	298
Table 7. Preparing Dilutions of Antimicrobial Agents to Be Used in Agar Dilution Susceptibility Tests .....	304
Table 8A. Preparing Dilutions of Antimicrobial Agents to Be Used in Broth Dilution Susceptibility Tests .....	306
Table 8B. Preparing Dilutions of Water-Insoluble Antimicrobial Agents to Be Used in Broth Dilution Susceptibility Tests .....	308
Appendix A. Suggestions for Confirming Antimicrobial Susceptibility Test Results and Organism Identification for Agents Approved by the US Food and Drug Administration for Clinical Use .....	310

## Contents (Continued)

Appendix B. Intrinsic Resistance .....	318
Appendix C. QC Strains for Antimicrobial Susceptibility Tests.....	324
Appendix D. Anaerobe Cumulative Antibiogram.....	330
Appendix E. Susceptible-Dose Dependent Interpretive Category .....	334
Appendix F. Epidemiological Cutoff Values .....	338
Appendix G. Using Molecular Assays for Resistance Detection .....	342
Appendix H. Cefiderocol Broth Preparation and Reading Broth Microdilution Minimal Inhibitory Concentration End Points .....	358
Glossary I (Part 1). $\beta$ -Lactams: Class and Subclass Designations and Generic Names .....	364
Glossary I (Part 2). Non- $\beta$ -Lactams: Class and Subclass Designations and Generic Names .....	368
Glossary II. Antimicrobial Agent Abbreviations, Routes of Administration, and Drug Class .....	372
Glossary III. List of Identical Abbreviations Used for More Than One Antimicrobial Agent in US Diagnostic Products .....	380
The Quality Management System Approach .....	382



## Overview of Changes

CLSI M100-Ed34 replaces the previous edition of the supplement, CLSI M100-Ed33, published in 2023. Major additions, reformatting, and/or table relocation changes are summarized below, followed by additional noteworthy changes detailed by section/table. Changes to content since the previous edition appear in boldface type; however, minor editorial or formatting changes are not listed here, nor highlighted in boldface type. To learn more about the organization of CLSI M100-Ed34, check the “Instructions for Use.”

CLSI M100 is updated and reviewed annually as new data and new agents become available. Use of outdated documents is strongly discouraged.

Major Additions/Revisions	
<ul style="list-style-type: none"> <li>• Tables 1 and Tables 2 (general): These tables were renumbered so that each Table 1 has a corresponding Table 2.</li> <li>• Table 1I: A new table for suggested drugs to test and report on <i>Neisseria meningitidis</i> was added.</li> <li>• Table 1J: A combined table for suggested drugs to test and report on anaerobes was added. Previously, these suggestions were listed in separate gram-negative and gram-positive tables.</li> <li>• Table 2A-2: A new table with breakpoints specific to <i>Salmonella</i> and <i>Shigella</i> spp. was added. Table 2A-1 no longer addresses these organism groups.</li> <li>• Tables 2, Tables 3, and former Appendix E: In previous editions of CLSI M100, dosage regimens were listed in Tables 2, 3E-2 (now 3F-2), 3E-3 (now 3F-3), and former Appendix E.                             <ul style="list-style-type: none"> <li>– Dosage regimens were removed from all Tables 2 and Tables 3F-2 and 3F-3.</li> <li>– Former Appendix E was reformatted, relocated to follow the Tables 2 containing breakpoints, and renamed “Table 2 Dosages. Antimicrobial Agent Dosage Regimens Used to Establish Susceptible or Susceptible-Dose Dependent Breakpoints” (referred to as “Table 2 Dosages” throughout).</li> </ul> </li> <li>• Tables 3 (general): These tables were renumbered to accommodate the addition of new Table 3D.</li> <li>• Table 3D: A new table describing a broth disk elution method for aztreonam plus ceftazidime-avibactam was added.</li> <li>• Table 3F-4: A new table with breakpoints specific to testing <i>Acinetobacter</i> spp. directly from positive blood cultures was added.</li> <li>• Table 3H: Former Tables 3G-1 and 3G-2, which described ancillary methods for testing oxacillin and cefoxitin against staphylococci, were condensed into Table 3H, which describes the oxacillin salt agar test only.</li> <li>• Appendixes (general): These sections were relabeled to accommodate the relocation of former Appendix E (now Table 2 Dosages).</li> </ul>	
Section/Table	Changes
General	
Throughout	<p><b>Added:</b></p> <ul style="list-style-type: none"> <li>• “Lower” qualifier to comment that daptomycin should not be routinely reported on organisms isolated from the lower respiratory tract</li> </ul> <p><b>Revised:</b></p> <ul style="list-style-type: none"> <li>• Suggestion for repeat testing of isolates initially susceptible that may develop resistance after initiation of therapy, from “within 3 to 4 days” to “within a few days”</li> </ul>

Overview of Changes (Continued)

Section/Table	Changes
<b>General (Continued)</b>	
<b>CLSI Breakpoint Additions Since 2010</b>	<p><b>Added:</b></p> <ul style="list-style-type: none"> <li>• Sulbactam-durlobactam disk diffusion and MIC breakpoints for <i>Acinetobacter</i> spp.</li> <li>• Tedizolid disk diffusion breakpoints for <i>Staphylococcus</i> spp. (<i>Staphylococcus aureus</i> only)</li> <li>• Tedizolid disk diffusion breakpoint for <i>Streptococcus</i> spp. <math>\beta</math>-hemolytic group (<i>Streptococcus pyogenes</i> and <i>Streptococcus agalactiae</i> only)</li> <li>• Tedizolid disk diffusion breakpoint for <i>Streptococcus</i> spp. viridans group (<i>Streptococcus anginosus</i> group only)</li> </ul>
<b>CLSI Breakpoint Revisions Since 2010</b>	<p><b>Revised:</b></p> <ul style="list-style-type: none"> <li>• Minocycline disk diffusion and MIC breakpoints for <i>Stenotrophomonas maltophilia</i></li> <li>• Linezolid disk diffusion breakpoints for <i>Staphylococcus</i> spp.</li> </ul> <p><b>Deleted:</b></p> <ul style="list-style-type: none"> <li>• Ceftazidime disk diffusion breakpoints for <i>Burkholderia cepacia</i> complex</li> <li>• Meropenem disk diffusion breakpoints for <i>B. cepacia</i> complex</li> <li>• Minocycline disk diffusion breakpoints for <i>B. cepacia</i> complex</li> <li>• Trimethoprim-sulfamethoxazole disk diffusion breakpoints for <i>B. cepacia</i> complex</li> <li>• Ceftazidime MIC breakpoints for <i>S. maltophilia</i></li> </ul>
<b>CLSI Archived Resources</b>	<p><b>Added:</b></p> <ul style="list-style-type: none"> <li>• Breakpoints that have been eliminated from CLSI M100 (detailed in CLSI Breakpoint Revisions Since 2010)</li> <li>• Test for Detecting Methicillin (Oxacillin) Resistance in <i>Staphylococcus</i> spp. table content related to detection of <i>mecA</i>-mediated resistance using cefoxitin or oxacillin (former Tables 3G-1 and 3G-2, revised to Table 3H)</li> <li>• QC range that has been eliminated from CLSI M100</li> </ul>
<b>CLSI Reference Methods vs Commercial Methods and CLSI vs US Food and Drug Administration Breakpoints</b>	<p><b>Deleted:</b></p> <ul style="list-style-type: none"> <li>• Final paragraph that referenced verification of breakpoints</li> </ul> <p><b>NOTE:</b> CLSI now provides or is in the process of updating and developing additional documents for validation and verification of susceptibility breakpoints.</p>
<b>Tables 1. Antimicrobial Agents That Should Be Considered for Testing and Reporting by Microbiology Laboratories</b>	
<b>Table 1B-2. <i>Acinetobacter</i> spp.</b>	<p><b>Added:</b></p> <ul style="list-style-type: none"> <li>• Sulbactam-durlobactam to Tier 3</li> </ul>
<b>Table 1B-4. <i>Stenotrophomonas maltophilia</i></b>	<p><b>Deleted:</b></p> <ul style="list-style-type: none"> <li>• Ceftazidime</li> </ul>

Overview of Changes (Continued)

Section/Table	Changes
<b>Tables 1. (Continued)</b>	
<b>Table 1D. <i>Enterococcus</i> spp.</b>	<b>Revised:</b> <ul style="list-style-type: none"> <li>Footnote d regarding susceptibility to penicillin</li> </ul>
<b>Table 1H-1. <i>Streptococcus</i> spp. <math>\beta</math>-Hemolytic Group</b>	<b>Added:</b> <ul style="list-style-type: none"> <li>Reference corresponding to intrapartum prophylaxis recommendations</li> </ul> <b>Revised:</b> <ul style="list-style-type: none"> <li>Footnote b regarding intrapartum prophylaxis recommendations</li> </ul>
<b>Table 1I. <i>Neisseria meningitidis</i></b>	New table
<b>Table 1J. Anaerobes</b>	New combined table <b>Added:</b> <ul style="list-style-type: none"> <li>Footnote a regarding tier placement of ampicillin and penicillin for anaerobes</li> </ul>
<b>Tables 2. Zone Diameter and/or MIC Breakpoints</b>	
<b>Introduction to Tables 2A–2J. Zone Diameter and MIC Breakpoints</b>	<b>Added:</b> <ul style="list-style-type: none"> <li>Introductory text for Tables 2A–2J and Table 2 Dosages</li> </ul>
<b>Table 2A-1. Zone Diameter and MIC Breakpoints for Enterobacterales (excluding <i>Salmonella/Shigella</i>)</b>	<b>Added:</b> <ul style="list-style-type: none"> <li>Comment regarding meropenem-vaborbactam and Enterobacterales that harbor OXA-48</li> <li>Comment to clarify suggested action when a carbapenemase marker is detected in an Enterobacterales isolate that is cefepime S or SDD</li> </ul>
<b>Table 2A-2. Zone Diameter and MIC Breakpoints for <i>Salmonella</i> and <i>Shigella</i> spp.</b>	New table
<b>Table 2B-2. Zone Diameter and MIC Breakpoints for <i>Acinetobacter</i> spp.</b>	<b>Added:</b> <ul style="list-style-type: none"> <li>General comment regarding using positive blood culture broth as an inoculum for direct disk diffusion testing</li> <li>Sulbactam-durlobactam disk diffusion and MIC breakpoints</li> </ul>
<b>Table 2B-3. MIC Breakpoints for <i>Burkholderia cepacia</i> complex</b>	<b>Deleted:</b> <ul style="list-style-type: none"> <li>General disk diffusion testing recommendations (disk diffusion no longer recommended for <i>B. cepacia</i>)</li> <li>Ceftazidime disk diffusion breakpoints</li> <li>Meropenem disk diffusion breakpoints</li> <li>Minocycline disk diffusion breakpoints</li> <li>Trimethoprim-sulfamethoxazole disk diffusion breakpoints</li> </ul>

Overview of Changes (Continued)

Section/Table	Changes
Tables 2. (Continued)	
<p><b>Table 2B-4. Zone Diameter and MIC Breakpoints for <i>Stenotrophomonas maltophilia</i></b></p>	<p><b>Added:</b></p> <ul style="list-style-type: none"> <li>• Comment regarding trimethoprim-sulfamethoxazole antimicrobial therapy</li> </ul> <p><b>Revised:</b></p> <ul style="list-style-type: none"> <li>• Minocycline disk diffusion and MIC breakpoints</li> </ul> <p><b>Deleted:</b></p> <ul style="list-style-type: none"> <li>• Ceftazidime MIC breakpoints</li> </ul>
<p><b>Table 2C. Zone Diameter and MIC Breakpoints for <i>Staphylococcus</i> spp.</b></p>	<p><b>Added:</b></p> <ul style="list-style-type: none"> <li>• Tedizolid disk diffusion breakpoints (<i>S. aureus</i>)</li> </ul> <p><b>Revised:</b></p> <ul style="list-style-type: none"> <li>• Linezolid disk diffusion breakpoints</li> <li>• Table describing methods or targets for detection of methicillin (oxacillin)-resistant <i>Staphylococcus</i> spp.</li> <li>• Text explaining <i>mecA</i>, ceftaxitin, and oxacillin and associated testing relationships</li> </ul> <p><b>Deleted:</b></p> <ul style="list-style-type: none"> <li>• Comment regarding MIC confirmation requirement for staphylococci resistant to linezolid by disk diffusion and requirement to read disk diffusion zones using transmitted light</li> </ul>
<p><b>Table 2H-1. Zone Diameter and MIC Breakpoints for <i>Streptococcus</i> spp. <math>\beta</math>-Hemolytic Group</b></p>	<p><b>Added:</b></p> <ul style="list-style-type: none"> <li>• Reference corresponding to intrapartum prophylaxis recommendations</li> <li>• Tedizolid disk diffusion breakpoint (<i>S. pyogenes</i> and <i>S. agalactiae</i> only)</li> </ul> <p><b>Revised:</b></p> <ul style="list-style-type: none"> <li>• Comment regarding intrapartum prophylaxis recommendations</li> </ul>
<p><b>Table 2H-2. Zone Diameter and MIC Breakpoints for <i>Streptococcus</i> spp. Viridans Group</b></p>	<p><b>Added:</b></p> <ul style="list-style-type: none"> <li>• Tedizolid disk diffusion breakpoint (<i>S. anginosus</i> group only)</li> </ul>
<p><b>Table 2 Dosages. Antimicrobial Agent Dosage Regimens Used to Establish Susceptible or Susceptible-Dose Dependent Breakpoints</b></p>	<p>New table (referred to as “Table 2 Dosages” throughout)</p> <p><b>Added:</b></p> <ul style="list-style-type: none"> <li>• Dosage for sulbactam-durlobactam for <i>Acinetobacter</i> spp.</li> <li>• Dosage for minocycline for <i>S. maltophilia</i></li> <li>• Dosage for ceftriaxone for MSSA</li> </ul>

Overview of Changes (Continued)

Section/Table	Changes
<b>Tables 3. Specialized Resistance Testing</b>	
<b>Table 3B. CarbaNP Test for Suspected Carbapenemase Production in Enterobacterales and <i>Pseudomonas aeruginosa</i></b>	<p><b>Added:</b></p> <ul style="list-style-type: none"> <li>• Reference pertaining to performance of the test</li> </ul>
<b>Table 3C. Modified Carbapenem Inactivation Methods for Suspected Carbapenemase Production in Enterobacterales and <i>Pseudomonas aeruginosa</i></b>	<p><b>Revised:</b></p> <ul style="list-style-type: none"> <li>• Test interpretation criterion from positive, negative, “indeterminate” to positive, negative, “inconclusive”</li> </ul>
<b>Table 3D. Aztreonam Plus Ceftazidime-Avibactam Broth Disk Elution Method</b>	New table
<b>Table 3F-1. Test for Performing Disk Diffusion Directly From Positive Blood Culture Broth</b>	<p><b>Added:</b></p> <ul style="list-style-type: none"> <li>• Breakpoint Additions/Revisions Since 2021 table for disk diffusion directly from positive blood culture broth</li> </ul> <p><b>Revised:</b></p> <ul style="list-style-type: none"> <li>• Table to include testing <i>Acinetobacter</i> spp. directly from positive blood cultures</li> </ul>
<b>Table 3F-2. Zone Diameter Disk Diffusion Breakpoints for Enterobacterales Direct From Blood Culture</b>	<p><b>Added:</b></p> <ul style="list-style-type: none"> <li>• Tobramycin 8-10 hour and 16-18 hour breakpoints</li> </ul> <p><b>Revised:</b></p> <ul style="list-style-type: none"> <li>• General comment regarding aztreonam, ceftazidime, and tobramycin breakpoints</li> </ul> <p><b>Deleted:</b></p> <ul style="list-style-type: none"> <li>• SDD column in table</li> </ul>
<b>Table 3F-3. Zone Diameter Disk Diffusion Breakpoints for <i>Pseudomonas aeruginosa</i> Direct From Blood Culture</b>	<p><b>Added:</b></p> <ul style="list-style-type: none"> <li>• Cefepime 16-18 hour and tobramycin 8-10 hour and 16-18 hour breakpoints</li> <li>• Comment regarding cefepime confirmatory MIC testing</li> </ul> <p><b>Deleted:</b></p> <ul style="list-style-type: none"> <li>• SDD column in table</li> </ul>
<b>Table 3F-4. Zone Diameter Disk Diffusion Breakpoints for <i>Acinetobacter</i> spp. Direct From Blood Culture</b>	New table
<b>Table 3H. Oxacillin Salt Agar Test for Detecting Methicillin (Oxacillin) Resistance in <i>Staphylococcus aureus</i></b>	<p><b>Deleted:</b></p> <ul style="list-style-type: none"> <li>• Content related to routine disk diffusion and MIC testing</li> </ul>

## Overview of Changes (Continued)

Section/Table	Changes
<b>Tables 3. (Continued)</b>	
<b>Table 3J. Tests for Detecting Inducible Clindamycin Resistance in <i>Staphylococcus</i> spp., <i>Streptococcus pneumoniae</i>, and <i>Streptococcus</i> spp. <math>\beta</math>-Hemolytic Group</b>	<p><b>Added:</b></p> <ul style="list-style-type: none"> <li>Reference corresponding to intrapartum prophylaxis recommendations</li> </ul> <p><b>Revised:</b></p> <ul style="list-style-type: none"> <li>Footnote b regarding intrapartum prophylaxis recommendations</li> <li>Reference pertaining to prevention of perinatal group B streptococcal disease updated to the American College of Obstetricians and Gynecologists guidelines</li> </ul>
<b>Tables 4. Disk Diffusion QC Ranges and Associated Tables</b>	
<b>Table 4A-1. Disk Diffusion QC Ranges for Nonfastidious Organisms and Antimicrobial Agents Excluding <math>\beta</math>-Lactam Combination Agents</b>	<p><b>Added:</b></p> <ul style="list-style-type: none"> <li>Footnote c regarding <i>S. aureus</i> ATCC<sup>®</sup> 43300 as a supplemental QC strain</li> </ul> <p><b>Revised:</b></p> <ul style="list-style-type: none"> <li>Linezolid QC range for <i>S. aureus</i> ATCC<sup>®</sup> 25923</li> <li>Tedizolid QC range for <i>S. aureus</i> ATCC<sup>®</sup> 25923</li> </ul> <p><b>Deleted:</b></p> <ul style="list-style-type: none"> <li>Footnote regarding reading zones of inhibition for linezolid and tedizolid for <i>S. aureus</i> ATCC<sup>®</sup> 25923 using transmitted light</li> </ul>
<b>Table 4A-2. Disk Diffusion QC Ranges for Nonfastidious Organisms and <math>\beta</math>-Lactam Combination Agents</b>	<p><b>Added:</b></p> <ul style="list-style-type: none"> <li>Column headers to highlight QC organisms recommended for routine QC testing</li> </ul>
<b>Tables 5. MIC QC Ranges and Associated Tables</b>	
<b>Table 5A-1. MIC QC Ranges for Nonfastidious Organisms and Antimicrobial Agents Excluding <math>\beta</math>-Lactam Combination Agents</b>	<p><b>Added:</b></p> <ul style="list-style-type: none"> <li>Footnote e regarding <i>S. aureus</i> ATCC<sup>®</sup> 43300 as a supplemental QC strain</li> <li>Footnote l regarding colistin QC organism alternatives to <i>P. aeruginosa</i> ATCC<sup>®</sup> 27853</li> <li>Footnote u regarding polymyxin B QC range for <i>Escherichia coli</i> NCTC 13846</li> <li>Upleganan QC ranges for <i>E. coli</i> ATCC<sup>®</sup> 25922 and <i>P. aeruginosa</i> ATCC<sup>®</sup> 27853</li> </ul> <p><b>Revised:</b></p> <ul style="list-style-type: none"> <li>Aztreonam QC range for <i>E. coli</i> ATCC<sup>®</sup> 25922</li> <li>Colistin QC range for <i>P. aeruginosa</i> ATCC<sup>®</sup> 27853</li> <li>Footnote o and associated figures regarding exebacase QC range for <i>S. aureus</i> ATCC<sup>®</sup> 29213 and additional testing guidance</li> </ul> <p><b>Deleted:</b></p> <ul style="list-style-type: none"> <li>Colistin QC range for <i>E. coli</i> ATCC<sup>®</sup> 25922</li> </ul>

Overview of Changes (Continued)

Section/Table	Changes
<b>Tables 5. (Continued)</b>	
<b>Table 5A-2. MIC QC Ranges for Nonfastidious Organisms and <math>\beta</math>-Lactam Combination Agents</b>	<p><b>Added:</b></p> <ul style="list-style-type: none"> <li>• Column headers to highlight QC organisms recommended for routine QC testing</li> <li>• Imipenem-funobactam QC ranges for:                             <ul style="list-style-type: none"> <li>– <i>E. coli</i> ATCC® 25922</li> <li>– <i>P. aeruginosa</i> ATCC® 27853</li> <li>– <i>Klebsiella pneumoniae</i> ATCC® 700603</li> <li>– <i>K. pneumoniae</i> ATCC® BAA-1705™</li> </ul> </li> </ul> <p><b>Revised:</b></p> <ul style="list-style-type: none"> <li>• Aztreonam QC ranges for:                             <ul style="list-style-type: none"> <li>– <i>E. coli</i> ATCC® 25922</li> <li>– <i>K. pneumoniae</i> ATCC® 700603</li> </ul> </li> </ul>
<b>Tables 6. Preparing Antimicrobial Agent Stock Solutions</b>	
<b>Table 6A. Solvents and Diluents for Preparing Stock Solutions of Antimicrobial Agents</b>	<p><b>Added:</b></p> <ul style="list-style-type: none"> <li>• Funobactam</li> <li>• Upleganan</li> </ul>
<b>Table 6C. Preparing Solutions and Media Containing Combinations of Antimicrobial Agents</b>	<p><b>Added:</b></p> <ul style="list-style-type: none"> <li>• Imipenem-funobactam</li> </ul>
<b>Appendixes</b>	
<b>Appendix B. Intrinsic Resistance, B1. Enterobacterales</b>	<p><b>Deleted:</b></p> <ul style="list-style-type: none"> <li>• Footnote g regarding <i>Serratia marcescens</i> and elevated MICs to tobramycin</li> </ul>
<b>Appendix C. QC Strains for Antimicrobial Susceptibility Tests</b>	<p><b>Added:</b></p> <ul style="list-style-type: none"> <li>• <i>E. coli</i> AR Bank #0348</li> <li>• Oxacillin MIC testing for MIC tests for <i>S. aureus</i> ATCC® 43300</li> </ul>
<b>Appendix G. Using Molecular Assays for Resistance Detection</b>	<p><b>Revised:</b></p> <ul style="list-style-type: none"> <li>• Table column headers</li> </ul>
<b>Table G3. Reporting Results From ESBL Resistance and Carbapenemase Molecular Tests for Enterobacterales</b>	<p><b>Added:</b></p> <ul style="list-style-type: none"> <li>• Text to clarify suggested action when a carbapenemase marker is detected in an Enterobacterales isolate that is cefepime S or SDD</li> </ul>

## Overview of Changes (Continued)

Section/Table	Changes
<b>Appendixes (Continued)</b>	
<b>Appendix H. Cefiderocol Broth Preparation and Reading Broth Microdilution Minimal Inhibitory Concentration End Points, H3. Determining Broth Microdilution End Points</b>	<p><b>Added:</b></p> <ul style="list-style-type: none"> <li>Figures showing determination of broth microdilution end points for cefiderocol</li> </ul> <p><b>Revised:</b></p> <ul style="list-style-type: none"> <li>Steps for reading and interpreting broth microdilution end points for cefiderocol, including figures showing determination of end points</li> </ul>
<b>Glossaries</b>	
<b>Glossary I (Part 1). <math>\beta</math>-Lactams: Class and Subclass Designations and Generic Names</b>	<p><b>Added:</b></p> <ul style="list-style-type: none"> <li>Imipenem-funobactam</li> </ul>
<b>Glossary I (Part 2). Non <math>\beta</math>-Lactams: Class and Subclass Designations and Generic Names</b>	<p><b>Added:</b></p> <ul style="list-style-type: none"> <li>Upleganan</li> </ul>
<b>Glossary II. Antimicrobial Agent Abbreviations, Routes of Administration, and Drug Class</b>	<p><b>Added:</b></p> <ul style="list-style-type: none"> <li>Imipenem-funobactam</li> <li>Upleganan</li> </ul>

Abbreviations: **AR**, antimicrobial resistance; ATCC<sup>®</sup>, American Type Culture Collection; MIC, minimal inhibitory concentration; **MSSA**, methicillin (oxacillin) susceptible *Staphylococcus aureus*; NCTC, National Collection of Type Cultures; QC, quality control; **S**, susceptible; **SDD**, susceptible-dose dependent.

### Footnote

a. ATCC<sup>®</sup> is a registered trademark of the American Type Culture Collection.



## Summary of CLSI Processes for Establishing Breakpoints and QC Ranges

The Clinical and Laboratory Standards Institute (CLSI) is an international, voluntary, not-for-profit, interdisciplinary, standards-developing, and educational organization accredited by the American National Standards Institute that develops and promotes the use of consensus-developed standards and guidelines within the health care community. These consensus standards and guidelines are developed in an open and consensus-seeking forum to cover critical areas of diagnostic testing and patient health care. CLSI is open to anyone or any organization that has an interest in diagnostic testing and patient care. Information about CLSI can be found at [www.clsi.org](http://www.clsi.org).

The CLSI Subcommittee on Antimicrobial Susceptibility Testing reviews data from a variety of sources and studies (eg, *in vitro*, pharmacokinetics/pharmacodynamics, and clinical studies) to establish antimicrobial susceptibility test methods, breakpoints, and QC parameters. The details of the data necessary to establish breakpoints, QC parameters, and how the data are presented for evaluation are described in CLSI M23.<sup>4</sup>

Over time, a microorganism's susceptibility to an antimicrobial agent may decrease, resulting in a lack of clinical efficacy and/or safety. In addition, microbiological methods and QC parameters may be refined to ensure more accurate and better performance of susceptibility test methods. Because of these types of changes, CLSI continually monitors and updates information in its documents. Although CLSI standards and guidelines are developed using the most current information available at the time, the field of science and medicine is always changing; therefore, standards and guidelines should be used in conjunction with clinical judgment, current knowledge, and clinically relevant laboratory test results to guide patient treatment.

Additional information, updates, and changes in this document are found in the meeting summary minutes of the CLSI Subcommittee on Antimicrobial Susceptibility Testing at <https://clsi.org/meetings/ast-file-resources/>.

## Instructions for Use of Tables

These instructions apply to:

- Tables 1A through 1J: suggested tiers of antimicrobial agents that should be considered for testing and reporting by microbiology laboratories. These suggestions include clinical efficacy, current consensus recommendations for first-choice and alternative drugs, and US Food and Drug Administration (FDA) clinical indications for use. In other countries, placement of antimicrobial agents in Tables 1A through 1J should be based on available drugs approved for clinical use by relevant regulatory organizations.
- Tables 2A through 2J: tables for each organism group that contain:
  - Recommended testing conditions
  - Routine QC recommendations (also see CLSI M02<sup>1</sup> and CLSI M07<sup>2</sup>)
  - General comments for testing the organism group and specific comments for testing particular agent/organism combinations
  - Agents that should be considered for routine testing and reporting by medical microbiology laboratories, as specified in Tables 1A through 1J (test/report Tiers 1, 2, 3, and 4), including agents reported only on organisms isolated from the urinary tract (designated by “U”)
  - Agents that are appropriate for the respective organism group but are not listed in Tables 1 and would generally not warrant routine testing by a medical microbiology laboratory in the United States (designated with an asterisk as “other”; designated with “Inv.” for “investigational” [not yet FDA approved]), including agents reported only on organisms isolated from the urinary tract (designated by “U”)
  - Zone diameter and minimal inhibitory concentration (MIC) breakpoints
- Tables 1J and 2J: tables containing specific recommendations for testing and reporting results on anaerobes and some of the information listed in the bullets above
- Tables 3A through 3L: tables describing tests to detect particular resistance types in specific organisms or organism groups

**Table 1B-1. *Pseudomonas aeruginosa***

Tier 1: Antimicrobial agents that are appropriate for routine, primary testing and reporting	Tier 2: Antimicrobial agents that are appropriate for routine, primary testing but may be reported following cascade reporting rules established at each institution	Tier 3: Antimicrobial agents that are appropriate for routine, primary testing in institutions that serve patients at high risk for MDROs but should only be reported following cascade reporting rules established at each institution.	Tier 4: Antimicrobial agents that may warrant testing and reporting by clinician request if antimicrobial agents in other tiers are not optimal because of various factors
Ceftazidime	Imipenem	Cefiderocol	
Cefepime	Meropenem	Ceftazidime-avibactam	
Piperacillin-tazobactam		Ceftolozane-tazobactam	
		Imipenem-relebactam	
Tobramycin			
Ciprofloxacin			
Levofloxacin			
			Aztreonam
<b>Urine Only</b>			
	Amikacin		

Abbreviation: MDRO, multidrug-resistant organism.

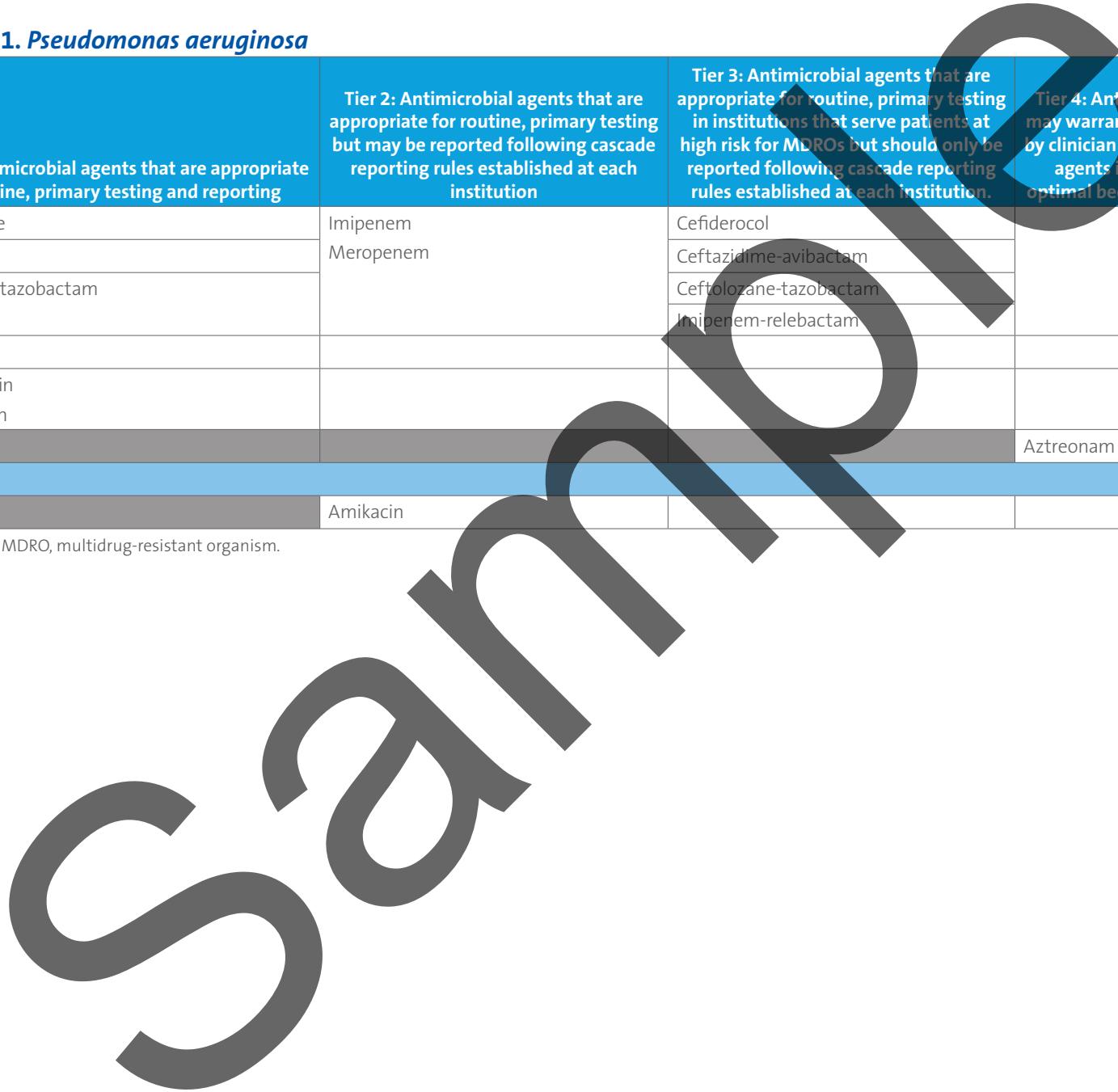


Table 1B-5. Other Non-Enterobacterales<sup>a,b</sup>

Tier 1: Antimicrobial agents that are appropriate for routine, primary testing and reporting	Tier 2: Antimicrobial agents that are appropriate for routine, primary testing but may be reported following cascade reporting rules established at each institution	Tier 3: Antimicrobial agents that are appropriate for routine, primary testing in institutions that serve patients at high risk for MDROs but should only be reported following cascade reporting rules established at each institution	Tier 4: Antimicrobial agents that may warrant testing and reporting by clinician request if antimicrobial agents in other tiers are not optimal because of various factors
Ceftazidime	Cefepime Imipenem Meropenem		
Gentamicin Tobramycin	Amikacin		
Piperacillin-tazobactam			
Trimethoprim-sulfamethoxazole			
	Aztreonam		
	Ciprofloxacin Levofloxacin		
	Minocycline		
			Cefotaxime Ceftriaxone
<b>Urine Only</b>			
Tetracycline <sup>c</sup>			

Abbreviations: MDRO, multidrug-resistant organism; MIC, minimal inhibitory concentration.

**Footnotes**

- a. Other non-Enterobacterales include *Pseudomonas* spp. and other nonfastidious, glucose-nonfermenting, gram-negative bacilli but exclude *Pseudomonas aeruginosa*, *Acinetobacter* spp., *Burkholderia cepacia* complex, and *Stenotrophomonas maltophilia*. Refer to each respective Table 1 for suggested antimicrobial agents to test and report.
- b. MIC testing only; disk diffusion test is unreliable.
- c. Organisms that are susceptible to tetracycline are also considered susceptible to doxycycline and minocycline. However, some organisms that are intermediate or resistant to tetracycline may be susceptible to doxycycline, minocycline, or both.

Table 1H-2. *Streptococcus* spp. Viridans Group

Tier 1: Antimicrobial agents that are appropriate for routine, primary testing and reporting	Tier 2: Antimicrobial agents that are appropriate for routine, primary testing but may be reported following cascade reporting rules established at each institution	Tier 3: Antimicrobial agents that are appropriate for routine, primary testing in institutions that serve patients at high risk for MDROs but should only be reported following cascade reporting rules established at each institution	Tier 4: Antimicrobial agents that may warrant testing and reporting by clinician request if antimicrobial agents in other tiers are not optimal because of various factors
Ampicillin <sup>a,b</sup> Penicillin <sup>a,b</sup>			
Cefotaxime Ceftriaxone			Cefepime
	Vancomycin		
		Linezolid Tedizolid <sup>c</sup>	
		Dalbavancin <sup>a,c</sup> Oritavancin <sup>a</sup> Telavancin <sup>a</sup>	
			Ceftolozane-tazobactam
			Clindamycin <sup>d</sup>
			Erythromycin <sup>d,e</sup>
			Levofloxacin

Abbreviations: MDRO, multidrug-resistant organism; MIC, minimal inhibitory concentration.

**Footnotes**

- a. MIC testing only; disk diffusion test is unreliable.
- b. **Rx:** Penicillin- or ampicillin-intermediate isolates may necessitate combined therapy with an aminoglycoside for bactericidal action.
- c. Report only on *S. anginosus* group (including *S. anginosus*, *S. intermedius*, and *S. constellatus*).
- d. Not routinely reported on organisms isolated from urinary tract.
- e. Susceptibility and resistance to azithromycin and clarithromycin can be predicted by testing erythromycin.

# Sample



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